

KREBS / HEME ASSIST (FGN - 011)

SUPPORT FOR HEALTHY HEME CYCLE AND KREBS CYCLE FUNCTION

FUNCTIONAL

KREBS/HEME ASSIST

Dietary Supplemen 90 Capsules

The Heme pathway in humans is involved in the production of a wide range of proteins involved in detoxification, antioxidant processes, gas delivery, electron transport, and signaling molecules. This makes the Heme pathway one of the most essential biochemical pathways among most living organisms.

The human Heme pathway is an intracellular, enzymatic process of eight sequential enzymatic reactions to produce heme. The intermediates of these enzymatic reactions are collectively called porphyrins and porphyrinogens, which are quickly converted from one intermediate to the next to the final product of heme. Heme itself acts as a regulatory molecule, tightly controlling the heme production process. The Heme pathway moves from the cytosol to the mitochondrion, back to the cytosol.⁵

The first step of the heme pathway is ALA synthase conversion, which occurs in the mitochondrion combining glycine from the mitochondrion amino acid pool and succinyl Co-A from the TCA cycle to produce δ -Aminolevulinic acid.^{5(p875)} In the second step of the heme pathway, the enzyme ALA dehydratase converts $\delta\text{-}$ Aminolevulinic acid to Porphobilinogen in the cytosol. The process continues in the cytosol in the next few steps as Porphobilinogen deaminase converts Porphobilinogen to Hydroxymethyl bilane in step three. In the fourth step, Uroporphyrinogen synthase converts Hydroxymethyl bilane to Uroporphyrinogen III. The fifth step involves the conversion of Uroporphyrinogen III to Coproporphyrinogen III by Uroporphyrinogen III decarboxylase. The pathway transitions back to the mitochondrion, with the conversion of Coproporphyrinogen III to Protoporphyrinogen IX by Coproporphyrinogen III oxidase in step six. Step seven converts Protoporphyrinogen IX to Protoporphyrin IX via the Protoporphyrinogen oxidase enzyme. The final step occurs in



the mitochondrion as the Ferrochelatase enzyme inserts Fe2+ into Protoporphyrin IX to produce heme.⁵

The following are all Heme-based proteins that occur in humans:

- Hemoglobin, Myoglobin, Neuroglobin, Cytoglobin involved in the delivery of oxygen to red blood cells, muscles, nerve cells, and brain, . respectively
- Cytochrome p450 – involved in Phase I detoxification
- Cytochrome B5 and Cytochrome c involved in the electron transport chain •
- Peroxidase and Catalase involved in hydrogen peroxide break down
- Tryptophan pyrrolase involved in the conversion of tryptophan to NAD and NADPH via the Kynurenine pathway .
- Nitric oxide synthase involved in production of nitric oxide from L-arginine . and BH4
- Sulfite Oxidase involved in conversion of sulfites to sulfates1-4 .

Without heme, these enzymes will not be produced. If these enzymes cannot be produced in adequate quantity due to lack of heme production, then downstream enzymatic supports will be fruitless.

Without Heme, these processes cannot occur properly and downstream upregulation support will fail. Heme Pathway Support Formula forms the foundation of supporting these mechanisms.

Heme/Krebs Assist contains the following nutrients to support function and is made with a vegetable capsule. 90 caps per bottle.

Heme Pathway Support formula provides necessary cofactors to optimize theheme pathway and the functioning of heme dependent processes:

Oxygen delivery to the tissues Phase I detoxification **Electron Transport** Hydrogen Peroxide degradation NADPH production Nitric Oxide production Sulfite to Sulfate conversion

Vitamin E (as d-Alpha Tocopheryl Succinate) may help reduce certain heme pathway dysregulation issues involved in skin symptoms^{6, 7} and oxidative stress.⁸

The heme pathway is initiated when succinyl-CoA is combined with glycine to form δ --aminolevulinic acid (ALA), a precursor to heme.⁹ The

succinate can be converted to succinyl-CoA in a reversible reaction by the enzyme Succinyl coenzyme A synthetase¹⁰ to support the initiation of the heme pathway.

L-Glycine is a cofactor along with Succinyl-Co-A at the top of the heme pathway to produce δ -aminolevulinic acid (ALA), a precursor to heme⁹

B3 (as Niacinamide) is essential for NAD+ production, which is a cofactor for the TCA cycle enzymes Pyruvate dehydrogenase complex, α -Ketoglutarate dehydrogenase complex, Isocitrate dehydrogenase, and Malate dehydrogenase.¹¹ B3 deficiency may contribute to increased heme dysregulation and porphyrin excretion.¹² Lack of B3 may affect NAD+ production and also impact glycine synthesis from serine.¹³

B2 (as Riboflavin 5 Phosphate) is a precursor of the coenzymes flavin mononucleotide and FAD needed in the TCA cycle for the enzymes pyruvate dehydrogenase, α -Ketoglutarate dehydrogenase complex, succinyl-CoA synthetase, and succinate dehydrogenase.¹¹ B2 is also a cofactor for protoporphyrinogen oxidase (PPOX) enzyme of the heme biosynthetic pathway.¹⁴ Lack of B2 can contribute to heme dysregulation and increased urinary coproporphyrins.¹⁵ B2 deficiency can also reduce B6 levels, another necessary cofactor in the TCA cycle and Heme pathway.¹⁶

Vitamin B6 (as Pyridoxal 5 Phosphate) is a necessary cofactor for the ALAS enzyme, a rate limiting step of heme synthesis.¹⁷, an important cofactor in the Kynurenine

Servings Per Container 30		
A	mount Per Serving	%D\
Vitamin E (as d-Alpha Tocopheryl Succinate)	50 IU	1679
Riboflavin (as Riboflavin 5 Phosphate)	15 mg	882
Niacin (as Niacinamide)	50 mg	250
Vitamin B-6 (as Pyridoxal 5 Phosphate)	25 mg	1250
Folate (as Quatrefolic L-5-Methyltetrahydrofolate, glucosamine sal	t) 200 mcg	50
Vitamin B12 (as Methylcobalamin)	25 mcg	
Biotin	1000 mcg	333
Zinc (as TRAACS® Zinc Bisglycinate Chelate)	10 mg	679
Thiamine (as Thiamine Mononitrate)	50 mg	3333
L-Glycine	100 mg	
L-Threonine	25 mg	
L-Isoleucine	25 mg	
L-Valine	25 mg	
Alpha Lipoic Acid	200 mg	
Vitamin C (as Ascorbyl Palmitate)	300 mg	
DiMagnesium Malate	200 mg	

Other ingredients: Micro Crystalline Cellulose (USP). Vegetable Capsule (cellulose, purified water).

No artificial colors, artificial flavors, milk or milk derivatives or sodium added.

pathway conversion of tryptophan to NAD+, which is a necessary cofactor for the TCA cycle.¹⁸ B6 is a necessary cofactor of the δ -Aminolevulinate synthase (ALAS) enzyme at the top of the heme pathway.¹⁹ B6 also serves as a possible activator of the Ferrochelatase (FECH) enzyme in the last step of the heme pathway.^{20, 21}

Vitamin C (as Ascorbyl Palmitate) – is a potent antioxidant that may reduce oxidative damage and restore expression of the heme enzyme Protoporphyrinogen oxidase PPOX.⁸ Vitamin C also supports the absorption and proper regulation of bioavailable iron,²² a necessary cofactor of the Ferrochelatase (FECH) enzyme at the last step of the Heme pathway.⁵

Magnesium (as Magnesium Malate) plays an important role in regulating iron,^{23, 24} which is a necessary cofactor in the last step of the heme pathway, the Ferrochelatase (FECH) enzyme.²⁵ Malate is an important intermediate of the TCA cycle.⁵

Folate (as Quatrefolic® - L-5-Methyltetrahydrofolate, glucosamine salt) is a non-essential activator of the Heme pathway enzyme Porphobilinogen-Deaminase (PBGD)²⁶ and may be involved in Uroporphyrinogen III Synthase (UROS) activity.²⁷ The active form of folate is also involved in glycine synthesis,²⁸ a required cofactor for the imitation of the Heme pathway.

Vitamin B12 (as Hydroxocobalamin) is a cofactor in the conversion of methylmalonyl-CoA to succinyl-CoA, the step in the TCA cycle immediately preceding the Heme Pathway.¹¹

Biotin is an essential cofactor for the metabolism of lipids, carbohydrates, and proteins needed to initiate the TCA cycle.²⁹, a cofactor for the TCA cycle enzymes pyruvate carboxylase, propionyl-CoA carboxylase, β -methylcrotonyl-CoA carboxylase and acetyl-CoA carboxylase,¹¹ which precedes the Heme pathway. Lack of biotin directly affects heme synthesis.³⁰

Zinc (as TRAACS® Zinc Bisglycinate Chelate) is required in the conversion of δ -Aminolevulinate (ALA) by the enzyme ALA dehydratase to Porphobilinogen³¹. Zinc deficiency may be a factor in Heme pathway dysregulation.³²

Thiamine (as Thiamine Mononitrate) is a cofactor for the pyruvate dehydrogenase enzyme responsible for pyruvate synthesis and for the α -ketoglutarate dehydrogenase enzyme required for succinyl-CoA synthesis, the step in the TCA cycle immediately preceding the Heme Pathway.¹¹

Lipoic Acid as Alpha Lipoic Acid is a required cofactor in the pyruvate dehydrogenase and α -Ketoglutarate dehydrogenase enzymes of the TCA cycle.¹¹ Alpha lipoic Acid may be a non-essential cofactor of the Uroporphyrinogen Decarboxylase (UROD) enzyme in the heme pathway.³³

L-Isoleucine, L-Threonine, and L-Valine are amino acids that are converted to Succinyl Co-A,¹¹ which is involved at the initiation of the heme pathway. Valine and isoleucine may be incorporated directly into δ -Aminolevulinic Acid via the enzyme ALA synthase (ALAS) more efficiently even than Succinyl Co-A.³⁴

References

1. Alderton WK, Cooper CE, Knowles RG. Nitric oxide synthases: structure, function and inhibition. *Biochemical Journal*. 2001;357(3):593-615. doi:10.1042/bj3570593.

2. Bonkovsky HL, Guo JT, Hou W, Li T, Narang T, Thapar M. Porphyrin and heme metabolism and the porphyrias. *Comprehensive Physiology*. 2013;3(1):365-401. doi:10.1002/cphy.c120006.

3. Paoli M, Marles-Wright J, Smith A. Structure–function relationships in heme-proteins. DNA and Cell Biology. 2002;21(4):271-280. doi:10.1089/104454902753759690.

4. Poulos TL. Heme enzyme structure and function. Chemical Reviews. 2014;114(7):3919-3962. doi:10.1021/cr400415k.

5. Lieberman M, Peet A. Marks' basic medical biochemistry: A clinical approach. 5th ed. Philadelphia: Wolters Kluwer Health; 2018.

6. Ayres S, Murphy G. Homozygous variegate porphyria: vitamin E as a possible therapeutic approach. *Journal of the Royal Society of Medicine*. 1987;80(2):129-129. doi:10.1177/014107688708000230.

7. Ayres JS, Mihan R. Porphyria cutanea tarda: response to vitamin E. A review and two case reports. Cutis. 1978;22(1):50-52.

8. Ferrer MD, Tauler P, Sureda A, Palacín C, Tur JA, Pons A. Variegate porphyria induces plasma and neutrophil oxidative stress: effects of dietary supplementation with vitamins E and C. *British Journal of Nutrition*. 2010;103(1):69-76. doi:10.1017/S0007114509991413.

9. Hunter GA, Ferreira GC. Molecular enzymology of 5-aminolevulinate synthase, the gatekeeper of heme biosynthesis. *Biochimica et Biophysica Acta* (*BBA*)-*Proteins and Proteomics*. 2011;1814(11):1467-1473. doi:10.1016/j.bbapap.2010.12.015.

10. Voet D, Voet JG. Biochemistry. 4th ed. NewYork: John Wiley & Sons; 2011.

11. Nelson DL, Lehninger AL, Cox MM. Lehninger principles of biochemistry. 5th ed. New York: Worth Publishers; 2008.

12. Cleary JP. The NAD deficiency diseases. Journal of Orthomolecular Medicine. 1986;1(3):149-157.

13. Shimizu Y, Sakuraba H, Doi K, Ohshima T. Molecular and functional characterization of D-3-phosphoglycerate dehydrogenase in the serine biosynthetic pathway of the hyperthermophilic archaeon Sulfolobus tokodaii. *Archives of Biochemistry and Biophysics*. 2008;470(2):120-128. doi:10.1016/j.abb.2007.11.010.

14. Layer G, Reichelt J, Jahn D, Heinz DW. Structure and function of enzymes in heme biosynthesis. *Protein Science*. 2010;19(6):1137-1161. doi:10.1002/pro.405.

15. Vannotti A. Porphyrins: Their Biological and Chemical Importance: Transl. London1954.

16. Sauberlich H. Interactions of thiamin, riboflavin, and other B-vitamins. *Annals of the New York Academy of Sciences*. 1980;355:80-97. doi: 10.1111/j.1749-6632.1980.tb21329.x.

17. Astner I, Schulze JO, Van den Heuvel J, Jahn D, Schubert WD, Heinz DW. Crystal structure of 5-aminolevulinate synthase, the first enzyme of heme biosynthesis, and its link to XLSA in humans. *The EMBO Journal*. 2005;24(18):3166-3177. doi:10.1038/sj.emboj.7600792.

18. Ueland PM, McCann A, Midttun Ø, Ulvik A. Inflammation, vitamin B6 and related pathways. *Molecular Aspects of Medicine*. 2017;53:10-27. doi:10.1016/j.mam.2016.08.001.

19. Turbeville TD. PLP-dependent [alpha]-oxoamine synthases: Phylogenetic analysis, structural plasticity, and structure-function studies on 5aminolevulinate synthase: University of South Florida; 2009.

20. Labbe RF, Nielsen L. Clinical-Biochemical Interpretations of Erythrocyte Protoporphyrin1. In: Freiburg S, editor. *Porphyrins in Human Diseases:* Karger Publishers; 1976. pp. 141-147.

21. de Molina Ríos M, de Catabbi Billi S, de Viale San LM. Liver ferrochelatase from normal and hexachlorobenzene porphyric rats. Mechanism of drug action. *The International Journal of Biochemistry*. 1991;23(7-8):669-673. doi: 10.1016/0020-711X(91)90036-M.

22. Lynch SR, Cook JD. Interaction of vitamin C and iron. Annals of the New York Academy of Sciences. 1980;355(3):33-44. doi:10.1111/j.1749-6632.1980.tb21325.x.

23. Kimura M, Yokoi K. Iron accumulation in tissues of magnesium-deficient rats with dietary iron overload. *Biological Trace Element Research*. 1996;51(2):177-197. doi:10.1007/BF02785437.

24. McDonald R, Keen CL. Iron, zinc and magnesium nutrition and athletic performance. Sports Medicine. 1988;5(3):171-184. doi:10.2165/00007256-198805030-00004.

25. Hunter GA, Al-Karadaghi S, Ferreira GC. Ferrochelatase: the convergence of the porphyrin biosynthesis and iron transport pathways. *Journal of Porphyrins and Phthalocyanines*. 2011;15(05n06):350-356. doi:10.1142/S108842461100332X.

26. Noriega GO, Juknat AA, Batlle AMdC. Non-essential activation of rat liver porphobilinogen-deaminase by folic acid. Zeitschrift für Naturforschung C: A Journal of Biosciences. 1992;47(5-6):416-419. doi:10.1515/znc-1992-0616.

27. Kohashi M, Clement RP, Tse J, Piper WN. Rat hepatic uroporphyrinogen III co-synthase. Purification and evidence for a bound folate coenzyme participating in the biosynthesis of uroporphyrinogen III. *Biochemical Journal*. 1984;220(3):755-765. doi:10.1042/bj2200755.

28. Rao NA, Ambili M, Jala VR, Subramanya H, Savithri H. Structure-function relationship in serine hydroxymethyltransferase. *Biochimica et Biophysica Acta (BBA)-Proteins and Proteomics*. 2003;1647(1-2):24-29. doi:10.1016/S1570-9639(03)00043-8.

29. Moss J, Lane MD. The biotin-dependent enzymes. Adv Enzymol Relat Areas Mol Biol. 1971;35:321-442.

30. Atamna H, Newberry J, Erlitzki R, Schultz CŚ, Ames BN. Biotin deficiency inhibits heme synthesis and impairs mitochondria in human lung fibroblasts. *The Journal of Nutrition*. 2007;137(1):25-30. doi:10.1093/jn/137.1.25.

31. Zhang L. Heme biology: the secret life of heme in regulating

diverse biological processes. Singapore: World Scientific Publishing Company; 2011.

32. Roman W. Zinc in porphyria. The American Journal of Clinical Nutrition. 1969;22(10):1290-1303. doi:10.1093/ajcn/22.10.1290.

33. Vilas GL, Aldonatti C, de Viale LSM, de Molina MdR. Effect of α lipoic acid amide on hexachlorobenzene porphyria. *IUBMB Life*. 1999;47(5):815-823. doi:10.1080/15216549900201903.

34. Cavender FL. The metabolic source of the succinyl-CoA molety of δ -aminolevulinic acid. *Biochemical Medicine*. 1971;5(6):515-520. doi:10.1016/0006-2944(71)90058-5.